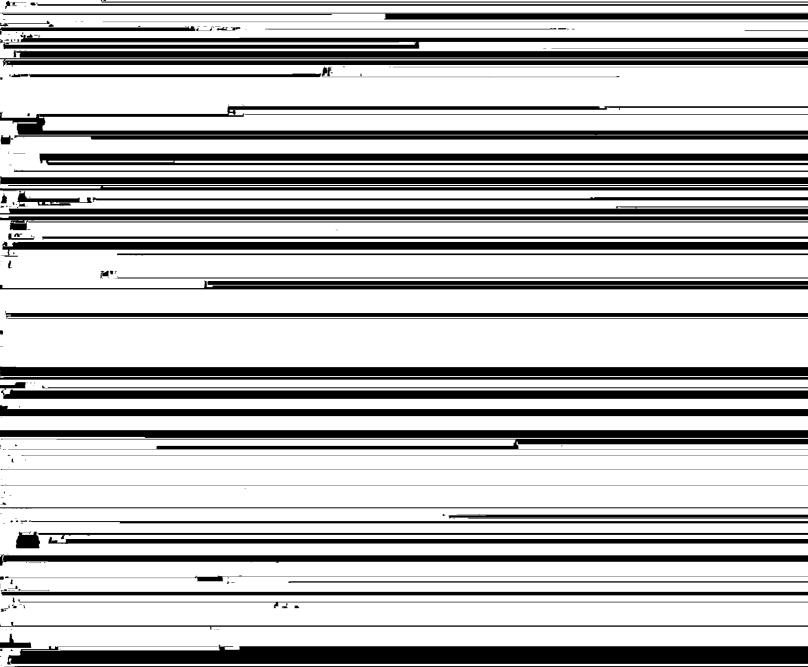
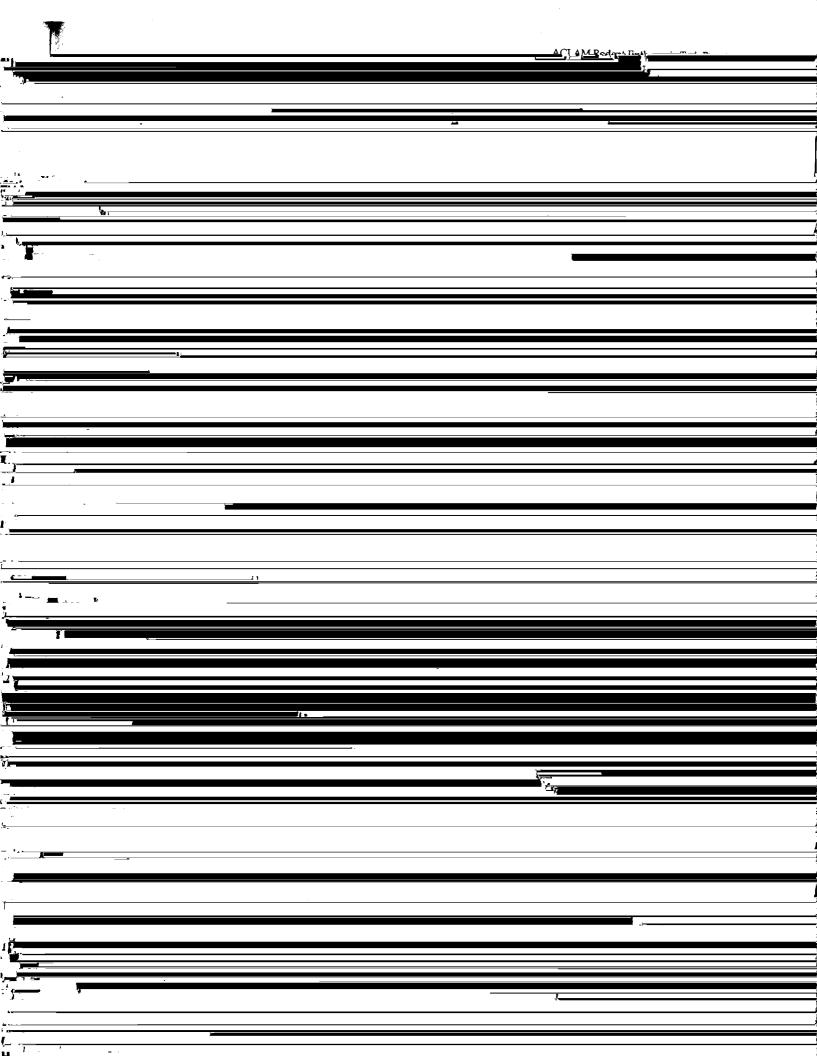
Public Statement

Report of the ACLAM Task Force on Rodent Euthanasia

James Artwohl, Patricia Brown, Brian Corning and Susan Stein

The ACLAM Task Force on Rodent Euthanasia was appointed by President Lynn Anderson in 2002 in response to growing concerns and controversy regarding techniques that were commonly used for rodent euthanasia. Three issues were targeted as the focus of the report: euthanasia of fetal and neonatal rodents, the use of carbon dioxide for rodent euthanasia, and the





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C. Euthanasia Chambers

- Euthanasia chambers should be kept clean and free of debris and excreta.
- The euthanasia chamber should be large enough to permit each animal to stand on the floor of the chamber with all 4 feet and have sufficient space to turn around and perform normal postural adjustments.

D. CO₂ Gas Delivery Systems

1. Sufficient carbon dioxide must be introduced into the chamber to totally displace the residual air by both mixing and dilution. Ideally, the inlet for delivery

increase distress for the animals. There is no conclusive evidence that adding pure oxygen to carbon dioxide makes this procedure less stressful to animals. ^{13,20,29,39} A fill rate of 20% of the chamber volume per minute with carbon dioxide, added to existing room air in the chamber should be appropriate to achieve a balanced gas mixture to fulfill the objective of rapid unconsciousness with minimal distress to the animals.

F. Cautionary Information

1. Animal carcasses should not be exposed to room air until

B. Biological Effects of Euthanasia Techniques

Table 1. Biologic effects of decapitation^{3,5,16,49,56,60,66}

Effect Mechanism

Increase in plasma sodium Increase in plasma potassium

Increase in GABA concentrations (brain)

Increase in Alanine (brain)

Increase in plasma ascorbic acid (30-40% > resting state)

Increase in blood catecholamine levels

Increased plasma calcium, magnesium No change in vasoactive intestinal peptides (brain)

No change in neuropeptide Y (brain)

Alteration in rat heart mitochondria function

Increase in serum corticosterone

Hemolysis

Continued postmortem neurochemical alterations

Stress stimulus \rightarrow mobilization from tissues to blood; generalized metabolic response secondary to sympathoadrenal response some handling related stimulation.

Possible handling stress

Table 2. Effects of physical and pharmacological euthanasia methods

		Table 4 Pialacia of American		
	Method of euthanasia	Table 4. Biologic effects of euthanasia induced by pharmacolog Effect	cic and/or physical methods	
	Injectable Pentobarbital ^{5,53,61}	Decreased muscular contractility in isolated	Mechanism	
		Decreased muscular contractility in isolated muscle preps Decreased GI smooth muscle contractility when given orally or intravenously; not seen in intraperitoneal goute.	Decreased calcium transport	
		given orally or intravenously; not seen in intraperitoneal		
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	Table 5. A postbotics, kotomics but 100 and
	Table 5. Anesthetics – ketamine hydrochloride, pentobarbital, chloral hydrate, chloralose and halothane in combination
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